



Pathogenicity Assessment of DNA Sequence Mutations

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2nd Practical Bioinformatics Course
Istanbul, 17/18 April 2017



**YOUR FUTURE
STARTS WITH HOPE**



Schedule

<i>Day 1</i>
Genome Biology in 2017
Bioinformatics Tools in Epigenetics
Pathogenicity Assessment of DNA Sequence Mutations
Introduction to Galaxy
Massive Data Sources
Practical

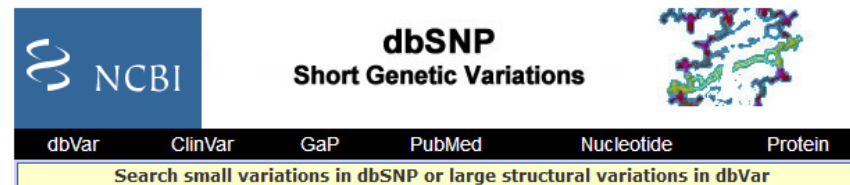
VCF Format

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1 166848563 . C G 100 PASS AA=C; AN=2184; ERATE=0.0009; VT=SNP; RSQ=0.8819; AC=33; LDAF=0.0166; SNPSOURCE=LOWCOV; THETA=0.0013; AVGPST=0.9949; AF=0.02; AMR_AF=0.06; AFR_AF=0.0020; EUR_AF=0.01
2 166903480 . C A 100 PASS AA=G; AN=2184; AC=6; THETA=0.0005; AVGPST=0.9996; VT=SNP; LDAF=0.0029; SNPSOURCE=LOWCOV; RSQ=0.9411; ERATE=0.0003; AF=0.0027; AFR_AF=0.01
3 166894356 . G A 100 PASS AC=297; LDAF=0.1395; RSQ=0.8949; AN=2184; THETA=0.0005; VT=SNP; AVGPST=0.9604; AA=A; ERATE=0.0000; SNPSOURCE=LOWCOV; AF=0.14; ASN_AF=0.01; AMR_AF=0.20; AFR_AF=0.03; EUR_AF=0.27
2 166198975 . G A 100 PASS ERATE=0.0004; AA=C; AN=2184; LDAF=0.0010; VT=SNP; AVGPST=0.9989; SNPSOURCE=LOWCOV; AC=1; THETA=0.0007; RSQ=0.5354; AF=0.0005; EUR_AF=0.0013
12 166852575 . G T 100 PASS ERATE=0.0004; RSQ=0.4926; THETA=0.0004; AA=G; AN=2184; AVGPST=0.9967; VT=SNP; LDAF=0.0022; SNPSOURCE=LOWCOV; AC=3; AF=0.0014; ASN_AF=0.0035; EUR_AF=0.0013
2 52062568 . G A 100 PASS ERATE=0.0005; RSQ=0.5636; AA=C; AN=2184; VT=SNP; THETA=0.0006; SNPSOURCE=LOWCOV; AC=3; AVGPST=0.9982; LDAF=0.0017; AF=0.0014; AFR_AF=0.01
4 52159534 . T A 100 PASS ERATE=0.0005; AA=C; AN=2184; AVGPST=0.9981; VT=SNP; LDAF=0.0004; SNPSOURCE=LOWCOV; AC=1; RSQ=0.4702; THETA=0.0003; AF=0.0005; AFR_AF=0.0020
9 130438189 . G A 100 PASS AA=C; AN=2184; RSQ=0.8976; LDAF=0.0046; VT=SNP; AC=10; AVGPST=0.9989; SNPSOURCE=LOWCOV; THETA=0.0010; ERATE=0.0003; AF=0.0046; AMR_AF=0.0028; AFR_AF=0.02
X 18606157 . G A 100 PASS ERATE=0.0004; AC=18; THETA=0.0004; AA=G; AN=2184; VT=SNP; LDAF=0.0087; RSQ=0.8246; SNPSOURCE=LOWCOV; AVGPST=0.9961; AF=0.01; AMR_AF=0.01; EUR_AF=0.02
9 130425622 . C T 100 PASS AA=C; AN=2184; AVGPST=0.9972; VT=SNP; AC=546; RSQ=0.9952; SNPSOURCE=LOWCOV; THETA=0.0007; LDAF=0.2496; ERATE=0.0007; AF=0.25; ASN_AF=0.17; AMR_AF=0.11; AFR_AF=0.73; EUR_AF=0.07
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8 133192493 . G A 100 PASS RSQ=0.9920; LDAF=0.2068; AA=G; AN=2184; AVGPST=0.9952; VT=SNP; THETA=0.0012; SNPSOURCE=LOWCOV; AC=452; ERATE=0.0006; AF=0.21; ASN_AF=0.02; AMR_AF=0.10; AFR_AF=0.63; EUR_AF=0.12
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11 33308577 . G A 100 PASS ERATE=0.0004; AA=G; RSQ=0.7293; AN=2184; AVGPST=0.9962; LDAF=0.0054; AC=8; VT=SNP; SNPSOURCE=LOWCOV; THETA=0.0010; AF=0.0037; ASN_AF=0.0017; EUR_AF=0.01
16 56385396 . T C 100 PASS AA=G; AN=2184; ERATE=0.0009; LDAF=0.2289; THETA=0.0008; VT=SNP; AC=501; AVGPST=0.9973; SNPSOURCE=LOWCOV; RSQ=0.9932; AF=0.23; ASN_AF=0.17; AMR_AF=0.16; AFR_AF=0.42; EUR_AF=0.18
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9 130982295 . G C 100 PASS RSQ=0.9108; THETA=0.0022; AA=G; AN=2184; AVGPST=0.9975; VT=SNP; LDAF=0.0107; SNPSOURCE=LOWCOV; AC=21; ERATE=0.0006; AF=0.01; AMR_AF=0.01; AFR_AF=0.0041; EUR_AF=0.02
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5 16792407 . C T 100 PASS LDAF=0.0016; AA=G; AN=2184; RSQ=0.6663; THETA=0.0005; VT=SNP; SNPSOURCE=LOWCOV; AVGPST=0.9987; AC=2; ERATE=0.0007; AF=0.0009; AMR_AF=0.0027; AFR_AF=0.0020
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10 89717712 . C T 100 PASS ERATE=0.0004; AN=2184; AC=6; VT=SNP; RSQ=0.8597; LDAF=0.0034; AA=A; SNPSOURCE=LOWCOV; THETA=0.0010; AVGPST=0.9987; AF=0.0027; AFR_AF=0.01
```

VCF Format from a SNP List

A list of up to 30,000 SNPs (rsIDs) can be converted to a VCF file (hg38 only) using this tool. A link to the VCF file is e-mailed when the conversion is complete.



The image shows the top section of the dbSNP website. On the left is the NCBI logo. In the center, the text 'dbSNP Short Genetic Variations' is displayed above a navigation bar with links for 'dbVar', 'ClinVar', 'GaP', 'PubMed', 'Nucleotide', and 'Protein'. To the right is a 3D molecular structure. Below the navigation bar is a search instruction: 'Search small variations in dbSNP or large structural variations in dbVar'.

The limit for each batch request is 30,000 IDs (ss#,rs#, etc.). A batch larger than rejected. However, there's no limit to the number of batches submitted.

Requested data will be sent to the email address you provide below.

(Please enter only one email address per request)

smithj@uni.edu

Organism

Homo sapiens

Enter RS Numbers

rs1800562
rs2395185
rs4813720

rs111
rs222
rs333
rs444
rs555

Example

Select Result Format

VCF

Submit

Whole Genome: dbWGFP

dbWGFP

A database and web server of human whole-genome single nucleotide variants and their functional predictions

JIANGLAB

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Introduction

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Introduction

The recent advancement of the next generation sequencing technology has enabled the fast and low-cost detection of all genetic variants spreading across entire human genomes, making the application of whole-genome sequencing a tendency in studies of disease-causing genetic variants. Nevertheless, there still lacks a repository that collects predictions of functionally damaging effects of human genetic variants, though it has been well recognized that such predictions plays a central role in the analysis of whole-genome sequencing data.

To fill in this gap, we developed **dbWGFP** (a database of human whole-genome single nucleotide variants and their functional predictions) that contains functional predictions and annotations of more than 8.5 billion possible human whole-genome single nucleotide variants. Specifically, this database integrates 32 functional predictions calculated by 13 popular computational methods, 15 conservation features derived from 4 conservation calculation approaches, and 44 valuable annotations obtained from the ENCODE project, accompanied with a highly efficient search program.

dbWGFP offers two web services for retrieving predictions and annotations for human whole-genome single nucleotide variants. In the [step-by-step mode](#), you can upload a file containing variants and retrieve results online. In the [batch mode](#), you can upload a file containing your email address and variants and retrieve results via your email.

dbWGFP offers two versions for downloading. The [lite version](#) includes prediction scores for human whole-genome single nucleotide variants. The [full version](#) includes both predictions and annotations. Both versions include a search program that can extract predictions and/or annotations in a highly efficient way. Different versions of dbWGFP are also [archived](#) for easy access.



Database, 2016, 1–11
doi: 10.1093/database/baw024
Original article



Original article

dbWGFP: a database and web server of human whole-genome single nucleotide variants and their functional predictions

Jiaxin Wu, Mengmeng Wu, Lianshuo Li, Zhuo Liu, Wanwen Zeng and Rui Jiang*

Whole Genome: dbWGFP

Table 1. Computational methods for predicting functionally damaging effects or conservation properties of single nucleotide variants

Method	Version	Source	Website
Grantham	Sep-74	CADD	—
SIFT	Aug-11	dbNSFP	http://sift.jcvi.org
PolyPhen-2	v2.2.2	dbNSFP	http://genetics.bwh.harvard.edu/pph2
LRT	Nov-09	dbNSFP	http://www.genetics.wustl.edu/jflab/lrt_query.html
MutationTaster	Mar-13	dbNSFP	http://www.mutationtaster.org
Mutation Assessor	Release 2	dbNSFP	http://mutationassessor.org
FATHMM	v2.3	dbNSFP	http://fathmm.biocompute.org.uk
RadialSVM	v2.4	dbNSFP	http://sites.google.com/site/jpopgen/dbNSFP
LR	v2.4	dbNSFP	http://sites.google.com/site/jpopgen/dbNSFP
CADD	v1.0	CADD	http://cadd.gs.washington.edu
GWAVA	v1.0	GWAVA	https://www.sanger.ac.uk/sanger/StatGen_Gwava
MSRV	Aug-07	MSRV	http://bioinfo.au.tsinghua.edu.cn/msrv
SinBaD	Nov-12	SinBaD	http://tingchenlab.cmb.usc.edu/sinbad
phastCons	Nov-09	UCSC	http://hgdownload.soe.ucsc.edu/goldenPath/hg19/phastCons46way
PhyloP	Nov-09	UCSC	http://hgdownload.soe.ucsc.edu/goldenPath/hg19/phyloP46way
GERP ++	May-11	GERP	http://mendel.stanford.edu/SidowLab/downloads/gerp
SiPhy	v0.5	SiPhy	http://www.broadinstitute.org/genome_bio/siphy

Whole Genome: gnomAD (SNP)

Variant: 6:26093141 G / A

	Exomes	Genomes	Total
Filter	Pass	Pass	
Allele Count	8157	1169	9326
Allele Number	246016	30926	276942
Allele Frequency	0.03316	0.03780	0.03367
dbSNP	rs1800562		
UCSC	6-26093141-G-A		
ClinVar	Click to search for variant in Clinvar		

Genotype Quality Metrics

Site Quality Metrics

Report this variant

Annotations

This variant falls on 14 transcripts in 1 genes:

missense

- HFE

Transcripts ▾

intron

- HFE - ENST00000352392

non coding transcript exon

- HFE

Transcripts ▾

Note: This list may not include additional transcripts in the same gene that the variant does not overlap.

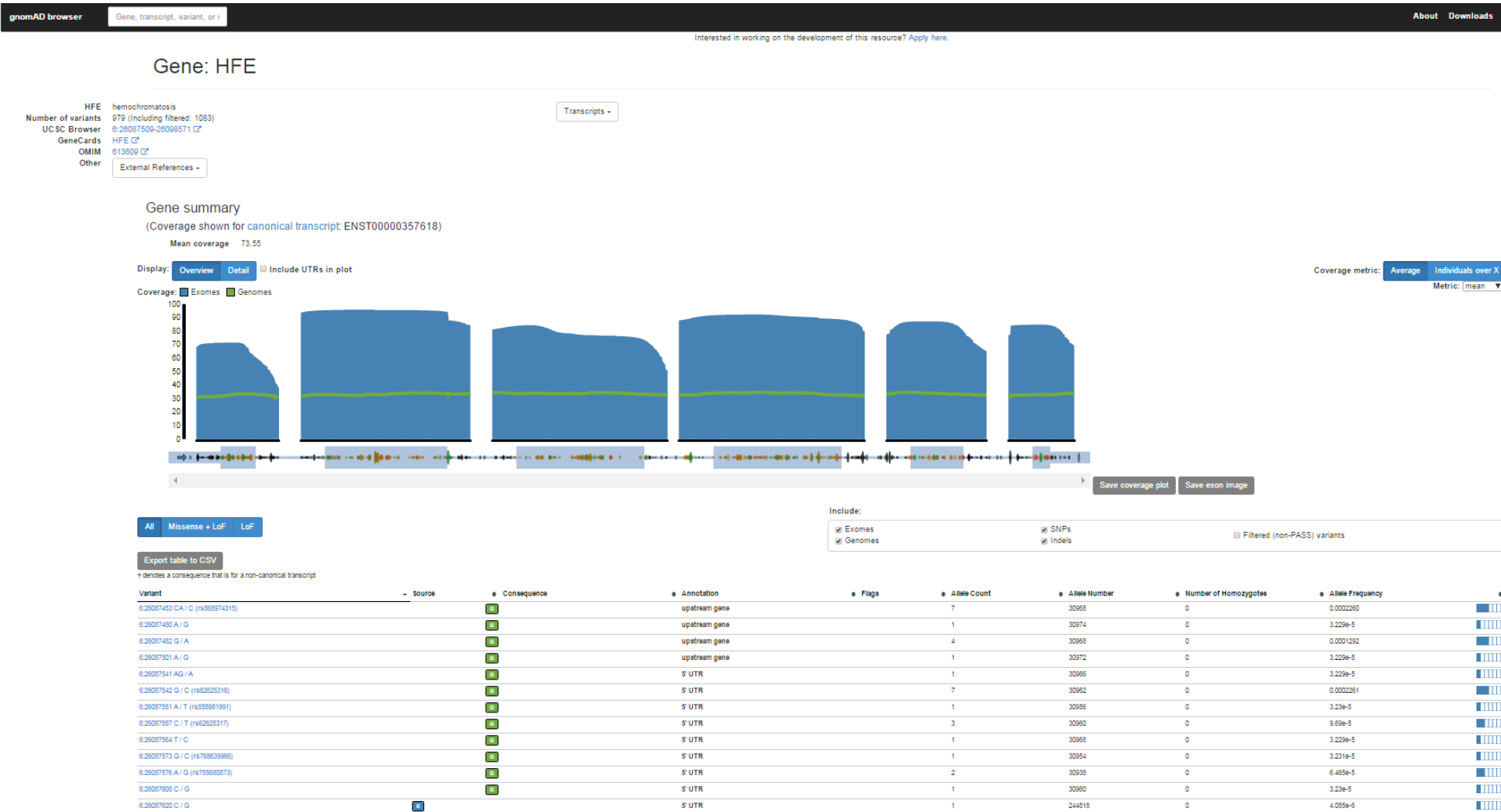
Population Frequencies

Population	Allele Count	Allele Number	Number of Homozygotes	Allele Frequency
European (Non-Finnish)	7275	126464	235	0.05753
Other	232	6464	5	0.03589
European (Finnish)	902	25778	20	0.03499
Latino	485	34414	5	0.01409
Ashkenazi Jewish*	119	10152	2	0.01172
African	241	24020	0	0.01003
South Asian	69	30782	0	0.002242
East Asian	3	18868	0	0.0001590
Total	9326	276942	267	0.03367

Include: ☒ Exomes ☒ Genomes

* For detailed analysis of Ashkenazi Jewish frequency see the [IBD Exomes Browser](#).

Whole Genome: gnomAD (gene)



Regulatory Variants



CellulAr dePendent dEactivating mutations predictor

Home

Examples

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Source Code

Flat files

Tissue

A549 EtOH 0.02pct Lung Carcin ▾

Model

eQTL
eQTL
dsQTL

SNP coordinate file

Browse...

No file selected.

(Maximum allowed file size 50.0 MB)

Paste in SNPs (dbSNP IDs or coordinates)

Coordinates format

dbSNP IDs format

Clear

View examples

[Examples](#)

Submit

Current Session's Jobs

- [Started at 11:20 a.m.](#)
- [Started at 3:49 p.m.](#)
- [Started at 11:33 a.m.](#)

Available data

Genomes

1. **Human (hg19)** (44 tissues)

Publications

1. Shan, L.; Vera, R.; Sharan, R.; Landsman, D. and Ovcharenko, I. [Quantifying deleterious effects of regulatory variants](#), *Nucleic Acids Research*. (2016) (DOI: 10.1093/nar/gkw1263):
2. Shan, L.; and Ovcharenko, I. [Human Enhancers Are Fragile and Prone to Deactivating Mutations](#), *Mol Biol Evol*. (2015) 32 (8):2161-2180

Published online 15 December 2016

Nucleic Acids Research, 2017, Vol. 45, No. 5 2307–2317
doi: 10.1093/nar/gkw1263

Quantifying deleterious effects of regulatory variants

Shan Li¹, Roberto Vera Alvarez¹, Roded Sharan², David Landsman¹ and Ivan Ovcharenko^{1,*}

¹Computational Biology Branch, National Center for Biotechnology Information, National Library of Medicine, National Institutes of Health, Bethesda, MD 20892, USA and ²School of Computer Science, Tel Aviv University, Tel Aviv 69978, Israel

FunSeq2 for Known and Unknown Variants



FunSeq2 - A flexible framework to prioritize regulatory mutations from cancer genome sequencing

Analysis

Results

Downloads

Documentation

FAQ

Whole Genome Query

Overview

This tool is specialized to prioritize somatic variants from cancer whole genome sequencing. It contains two components : 1) building data context from various resources; 2) variants prioritization. We provide downloadable scripts for users to customize the data context (found under 'Downloads'). The variants prioritization step is downloadable, and also implemented as a web server (Right Panel), along with the pre-processed data context.

Instructions

- ❖ Input File - BED or VCF formatted. Click the "green" button to add multiple files. With multiple files, the tool will do recurrent analysis. (Note: for BED format, user can put variants from multiple genomes into one file, see [Sample input file](#) .)
- ❖ Recurrence DB - User can select particular cancer types from the database. The DB will continue to be updated with newly-available WGS data.
- ❖ Gene List - Option to analyze variants associated with a particular set of genes. Note: Please use Gene Symbols, with one row per gene.
- ❖ Differential Gene Expression Analysis - Option to detect differentially expressed genes in RNA-Seq data. Two files are needed: an expression file and a class label file. Please refer to [Expression input files](#) for instructions on how to prepare these files.

❖ Note: In addition to on-site calculation, we also provide scores for all possible noncoding SNVs of GRCh37/hg19 under 'Downloads' (without annotation and recurrence analysis).

Input File: (only for hg19 SNVs)

No file chosen



BED or VCF files as input. [Sample input file](#)

Output Format:

bed ▼

MAF:

0

Minor allele frequency threshold to filter polymorphisms from 1KG (value 0~1)

Cancer Type from Recurrence DB: [Summary table](#)

All Cancer Types ▼

[Add a gene list](#) (Optional)

[Add differential gene expression analysis](#) (Optional)

Integrative Annotation of Variants from 1092 Humans: Application to Cancer Genomics

Ekta Khurana^{1,2,*}, Yao Fu^{1,*}, Vincenza Colonna^{3,4,*}, Xinmeng Jasmine Mu^{1,*}, Hyun Min Kang⁵, Tuuli Lappalainen^{6,7,8}, Andrea Sboner^{9,10}, Lucas Lochovsky¹, Jieming Chen¹¹, Arif Harmanci^{1,2}, Jishnu Das^{12,13}, Alexej Abyzov^{1,2}, Suganthi Balasubramanian^{1,2}, Kathryn Beal¹⁴, Dimple Chakravarty⁹, Daniel Challis¹⁵, Yuan Chen³, Declan Clarke¹⁶, Laura Clarke¹⁴, Fiona Cunningham¹⁴, Uday S. Evani¹⁹, Paul Flicek¹⁴, Robert Fragoza^{13,17}, Erik Garrison¹⁸, Richard Gibbs¹⁵, Zeynep H. Gümüş^{10,19}, Javier Herrero¹⁴, Naoki Kitabayashi⁹, Yong Kong^{2,20}, Kasper Lage^{21,22,23,24,25}, Vaja Liluashvili^{10,19}, Steven M. Lipkin²⁶, Daniel G. MacArthur^{22,27}, Gabor Marth¹⁸, Donna Muzny¹⁵, Tunc H. Pers^{24,28,29}, Graham R. S. Ritchie¹⁴, Jeffrey A. Rosenfeld^{30,31,32}, Cristina Sisu^{1,2}, Xiaomu Wei^{13,26}, Michael Wilson^{1,33}, Yali Xue³, Fuli Yu¹⁵, 1000 Genomes Project Consortium¹, Emmanouil T. Dermitzakis^{6,7,8}, Haiyuan Yu^{12,13}, Mark A. Rubin³, Chris Tyler-Smith^{3,4}, Mark Gerstein^{1,2,34,4}

RVS for Known and Unknown (MS) Variants

[RVS](#)[Queries](#)[API](#)[Data sources](#)[About](#)[Contact Us](#)[DIVAS](#)

the Reference Variant Store

The Reference Variant Store holds information on more than 520 million genetic variants using various methods of annotation. Sources for variants in RVS are re-sequencing projects such as the 1000 Genomes, ESP6500, UK10K, TCGA, Scripps Welllderly; clinical annotation databases such as ClinVar and HGMD; and hypothetical, amino-acid changing single-point mutations from dbNSFP. We provide annotations on low level effects, phenotypes and diseases, population frequencies, and predictive functional impact scores.

Available queries

Our query interface is available via the "Queries" menu. Entry points are searches by specific coordinate, dbSNP, or gene; or a comparison of cohorts based on populations in RVS. The first four queries are also wrapped in our [REST API](#).

Query by dbSNP: enter dbSNP IDs (rs-numbers) to search RVS for matching variants.

Query by coordinates: enter specific chromosomal locations with alternate alleles to search RVS for genetic variants; or enter chromosomal regions to search RVS for all variants falling into those regions.

Query by gene: enter one or more genes and, optionally, genetic variants to search RVS.

Query by phenotype: enter a phenotype or disease name, and optionally a gene to search.

Query by cohorts: choose two cohorts from the available datasets and find the differences in allele frequencies with respect to selected genes.

Hakenberg et al. *BMC Bioinformatics* (2016) 17:24
DOI 10.1186/s12859-015-0865-9

BMC Bioinformatics

DATABASE

Open Access



Integrating 400 million variants from 80,000 human samples with extensive annotations: towards a knowledge base to analyze disease cohorts

Jörg Hakenberg^{1,3*}, Wei-Yi Cheng^{1,4}, Philippe Thomas^{2,5}, Ying-Chih Wang¹, Andrew V. Uzilov¹ and Rong Chen^{1*}

[RVS](#)[Queries](#)[API](#)[Data sources](#)[About](#)[Contact Us](#)[DIVAS](#)

RVS queries

[Query by coordinates](#)[Query by dbSNP](#)[Query by gene list](#)[Query by phenotype](#)[Query by cohorts \(beta\)](#)

Paste a list of max. 10 variants using coordinates and ref/alt alleles. Alternatively, enter a chromosomal region to retrieve all variants found in that region.

Example for a single nucleotide variant: chr7:140453136A>T

Example for a chromosomal region: 7:140430000-140440000 — we accept regions of up to 10,000bp.

6:32400000-32450000



Limit: ☐ exonic ☐ non-synonymous ☒ all

☒ Display only variants observed in 1000KG Ph3, ExAC, ESP6500, UK10K ALSPAC/TWINS, and/or Scripps Welllderly

Output: ☐ Display results for the canonical isoform only (where applicable)

Format: ☒ HTML ☐ JSON ☐ Excel

Search

[Click here to paste examples](#)

ClinGen Pathogenicity Calculator

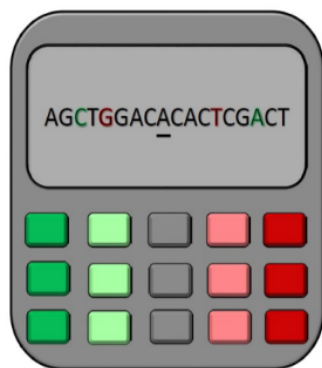


PATHOGENICITY CALCULATOR

Article describing ClinGen Pathogenicity Calculator is now available online from [Genome Medicine](#).

LOGIN

WHAT IS THE CLINGEN PATHOGENICITY CALCULATOR?



The shift from genetic testing of individual genes to exome and genome sequencing has been accompanied by new challenges in genome interpretation. The American College of Medical Genetics and Genomics and the Association for Molecular Pathology (ACMG/AMP) have published [Standards and Guidelines for the Interpretation of Sequence Variants](#). To enable wide application of the ACMG/AMP and similar guidelines and the development of collective knowledge by the community, ClinGen has developed the ClinGen Pathogenicity Calculator. By automating the formal reasoning, the Calculator eliminates errors in rule application and makes it possible to automatically calculate provisional conclusions based on latest evidence. Moreover, the Calculator makes reasoning explicit by documenting applicable rules, evidence codes, and links to supporting data. By explicitly communicating the reasoning behind a conclusion about pathogenicity of any specific variant, the Calculator enables critical evaluation of the reasoning and facilitates resolution of conflicting conclusions.

SOFTWARE

Open Access



ClinGen Pathogenicity Calculator: a configurable system for assessing pathogenicity of genetic variants

Ronak Y. Patel^{1*}, Neethu Shah^{1†}, Andrew R. Jackson¹, Rajarshi Ghosh¹, Piotr Pawliczek¹, Sameer Paithankar¹, Aaron Baker¹, Kevin Riehle¹, Hallin Chen¹, Sofia Milosavljevic¹, Chris Bizon², Shawn Rynearson³, Tristan Nelson⁴, Gail P. Jarvik⁵, Heidi L. Rehm^{5,6}, Steven M. Harrison⁵, Danielle Azzariti⁵, Bradford Powell⁷, Larry Babb⁸, Sharon E. Plon¹, Aleksandar Milosavljevic^{1*} and on behalf of the ClinGen Resource

Exonic Variants: Known SNPs

MSV3d

Full text search



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Database of human missense variants mapped to 3D protein structures & *Annotation pipeline*



Download [XML] [CSV] [Text]

	Gene	Protein	AA change	dbSNP	Phenotype	PDB	Chain	Detail	SIFT score	PolyPhen-2 score	KD4V prediction
1	HFE	Q30201	p.Cys282Tyr	rs1800562	Hemochromatosis (HFE) [MIM:235200]	1de4	G	Detail	0.00 (D)	1 (D)	D (Predicted by rules : 1)



Download [XML] [CSV] [Text]

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This site is optimised for Firefox.

<http://decryphon.igbmc.fr/msv3d/cgi-bin/humsavar?rsid=rs1800562>



ORIGINAL ARTICLE

Comparison and integration of deleteriousness prediction methods for nonsynonymous SNVs in whole exome sequencing studies

Chengliang Dong^{1,2,†}, Peng Wei^{4,6,†}, Xueqiu Jian⁵, Richard Gibbs⁷, Eric Boerwinkle^{4,5,7}, Kai Wang^{1,2,3,*} and Xiaoming Liu^{4,5,*}

Abstract

Accurate deleteriousness prediction for nonsynonymous variants is crucial for distinguishing pathogenic mutations from background polymorphisms in whole exome sequencing (WES) studies. Although many deleteriousness prediction methods have been developed, their prediction results are sometimes inconsistent with each other and their relative merits are still unclear in practical applications. To address these issues, we comprehensively evaluated the predictive performance of 18 current deleteriousness-scoring methods, including 11 function prediction scores (PolyPhen-2, SIFT, MutationTaster, Mutation Assessor, FATHMM, LRT, PANTHER, PhD-SNP, SNAP, SNPs&GO and MutPred), 3 conservation scores (GERP++, SiPhy and PhyloP) and 4 ensemble scores (CADD, PON-P, KGGSeq and CONDEL). We found that FATHMM and KGGSeq had the highest discriminative power among independent scores and ensemble scores, respectively. Moreover, to ensure unbiased performance evaluation of these prediction scores, we manually collected three distinct testing datasets, on which no current prediction scores were tuned. In addition, we developed two new ensemble scores that integrate nine independent scores and allele frequency. Our scores achieved the highest discriminative power compared with all the deleteriousness prediction scores tested and showed low false-positive prediction rate for benign yet rare nonsynonymous variants, which demonstrated the value of combining information from multiple orthologous approaches. Finally, to facilitate variant prioritization in WES studies, we have pre-computed our ensemble scores for 87 347 044 possible variants in the whole-exome and made them publicly available through the ANNOVAR software and the dbNSFP database.

dbNSFP

DATABASES

Human Mutation



dbNSFP v3.0: A One-Stop Database of Functional Predictions and Annotations for Human Nonsynonymous and Splice-Site SNVs

Xiaoming Liu,^{1,2*} Chunlei Wu,³ Chang Li,² and Eric Boerwinkle^{1,2,4}

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Jpopgen - a collection of tools for DNA sequence analyses
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About the maintainer

This site is maintained by:

Xiaoming Liu, Ph.D.

Assistant Professor,
Human Genetics Center,
School of Public Health,
The University of Texas Health
Science Center at Houston.

E-mail: xmliu.uth@tghmail.com

[Homepage](#)

dbNSFP

INTRODUCTION:

dbNSFP is a database developed for functional prediction and annotation of all potential non-synonymous single-nucleotide variants (nsSNVs) in the human genome. Its current version is based on the Gencode release 22 / Ensembl version 79 and includes a total of 83,422,341 nsSNVs and ssSNVs (splicing-site SNVs). It compiles prediction scores from 20 prediction algorithms (SIFT, Polyphen2-HDIV, Polyphen2-HVAR, LRT, MutationTaster2, MutationAssessor, FATHMM, MetaSVM, MetaLR, CADD, VEST3, PROVEAN, FATHMM-MKL coding, fitCons, DANN, GenoCanyon, Eigen coding, Eigen-PC, M-CAP, REVEL, MutPred), 6 conservation scores (PhyloP x 2, phastCons x 2, GERP++ and SiPhy) and other related information including allele frequencies observed in the 1000 Genomes Project phase 3 data, UK10K cohorts data, ExAC consortium data and the NHLBI Exome Sequencing Project ESP6500 data, various gene IDs from different databases, functional descriptions of genes, gene expression and gene interaction information, etc.

Some dbNSFP contents (may not be up-to-date though) can also be accessed through [variant tools](#), [ANNOVAR](#), [KGGSeq](#), UCSC Genome Browser's [Variant Annotation Integrator](#), [Ensembl Variant Effect Predictor](#), [SnpSift](#) and [HGMD](#). Please cite our papers (see below) if you used dbNSFP contents through those tools.

Please note some component score/content of dbNSFP has specific requirements or licence for non-academic usage. dbNSFP does not grant the non-academic usage of those scores/contents, so please contact the original score/content providers for that purpose.

Please join our [Email group](#) for news and updates from dbNSFP.

For whole genome annotation, we recommend our whole genome annotation pipeline [WGSA](#), in which dbNSFP is a component resource.

We thank Drs. CS (Jonathan) Liu from [Softgenetics](#) and Kai Wang from [USC](#) for providing hosting space.

We welcome developers of functional prediction methods to provide their predictions and scores to the database. Please contact Dr. Xiaoming Liu (xmliu.uth@tghmail.com).

CITATION:

1. Liu X, Jian X, and Boerwinkle E. 2011. [dbNSFP: a lightweight database of human non-synonymous SNPs and their functional predictions](#). *Human Mutation*. 32:894-899.
2. Liu X, Jian X, and Boerwinkle E. 2013. [dbNSFP v2.0: A Database of Human Non-synonymous SNVs and Their Functional Predictions and Annotations](#). *Human Mutation*. 34:E2393-E2402.
3. Liu X, Wu C, Li C and Boerwinkle E. 2016. [dbNSFP v3.0: A One-Stop Database of Functional Predictions and Annotations for Human Non-synonymous and Splice Site SNVs](#). *Human Mutation*. 37:235-241. [[preprint](#)]

dbNSFP

DATABASES

Human Mutation

dbNSFP v3.0: A One-Stop Database of Functional Predictions and Annotations for Human Nonsynonymous and Splice-Site SNVs



Xiaoming Liu,^{1,2*} Chunlei Wu,³ Chang Li,² and Eric Boerwinkle^{1,2,4}

Index of dbNSFP prediction files

Name	Size	Last Modified	
dbnsfp_chr1_mutpred.txt.gz	195 MB	8/31/16	4:55:00 PM
dbnsfp_chr2_mutpred.txt.gz	143 MB	8/31/16	4:56:00 PM
dbnsfp_chr3_mutpred.txt.gz	108 MB	8/31/16	4:57:00 PM
dbnsfp_chr4_mutpred.txt.gz	75 MB	8/31/16	4:58:00 PM
dbnsfp_chr5_mutpred.txt.gz	89 MB	8/31/16	4:58:00 PM
dbnsfp_chr6_mutpred.txt.gz	97 MB	8/31/16	4:59:00 PM
dbnsfp_chr7_mutpred.txt.gz	90 MB	8/31/16	5:00:00 PM
dbnsfp_chr8_mutpred.txt.gz	66 MB	8/31/16	5:00:00 PM
dbnsfp_chr9_mutpred.txt.gz	79 MB	8/31/16	5:01:00 PM
dbnsfp_chr10_mutpred.txt.gz	76 MB	8/31/16	4:48:00 PM
dbnsfp_chr11_mutpred.txt.gz	114 MB	8/31/16	4:49:00 PM
dbnsfp_chr12_mutpred.txt.gz	100 MB	8/31/16	4:50:00 PM
dbnsfp_chr13_mutpred.txt.gz	34 MB	8/31/16	4:50:00 PM
dbnsfp_chr14_mutpred.txt.gz	61 MB	8/31/16	4:50:00 PM
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dbnsfp_chr16_mutpred.txt.gz	79 MB	8/31/16	4:51:00 PM
dbnsfp_chr17_mutpred.txt.gz	106 MB	8/31/16	4:52:00 PM
dbnsfp_chr18_mutpred.txt.gz	30 MB	8/31/16	4:52:00 PM
dbnsfp_chr19_mutpred.txt.gz	122 MB	8/31/16	4:53:00 PM
dbnsfp_chr20_mutpred.txt.gz	47 MB	8/31/16	4:55:00 PM
dbnsfp_chr21_mutpred.txt.gz	20 MB	8/31/16	4:55:00 PM
dbnsfp_chr22_mutpred.txt.gz	41 MB	8/31/16	4:55:00 PM
dbnsfp_chrX_mutpred.txt.gz	72 MB	8/31/16	5:01:00 PM
dbnsfp_chrY_mutpred.txt.gz	566 KB	8/31/16	5:01:00 PM
README	4 KB	9/1/16	2:55:00 PM

dbNSFP v3.0: A One-Stop Database of Functional Predictions and Annotations for Human Nonsynonymous and Splice-Site SNVs

Xiaoming Liu,^{1,2*} Chunlei Wu,³ Chang Li,² and Eric Boerwinkle^{1,2,4}

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About the maintainer

This site is maintained by:

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Assistant Professor,
Human Genetics Center,
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E-mail: xmlu.uth[at]gmail.com

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For whole genome annotation, we recommend our whole genome annotation pipeline [WGSA](#), in which dbNSFP is a component resource.

- **WGSA:** WGSA is an annotation pipeline for human genome re-sequencing studies, to facilitate the functional annotation step of whole genome sequencing (WGS). Currently WGSA supports the annotation of SNVs and indels locally without remote database requests, allowing it to scale up for large WGS studies.

1. Liu X, White S, Peng B, Johnson AD, Brody JA, Li AH, Huang Z, Carroll A, Wei P, Gibbs R, Klein RJ and Boerwinkle E. 2016. **WGSA: an annotation pipeline for human genome sequencing studies.** *Journal of Medical Genetics* 53:111-112.

WGSA: <https://sites.google.com/site/jpopgen/wgsa>

WGSA



WGSA: an annotation pipeline for human genome sequencing studies

Xiaoming Liu, Simon White, Bo Peng, Andrew D Johnson, Jennifer A Brody, Alexander H Li, Zhuoyi Huang, Andrew Carroll, Peng Wei, Richard Gibbs, Robert J Klein and Eric Boerwinkle

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About the maintainer

This site is maintained by:

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Assistant Professor,
Human Genetics Center,
School of Public Health,
The University of Texas Health
Science Center at Houston.

E-mail: xmliu.uth@gmail.com

[Homepage](#)

WGS Annotator (WGSA) - an annotation pipeline for human genome re-sequencing studies

WGSA is an annotation pipeline for human genome re-sequencing studies, to facilitate the functional annotation step of whole genome sequencing (WGS). Currently WGSA supports the annotation of SNVs and indels locally without remote database requests, allowing it to scale up for large WGS studies.

For gene-model based annotation, WGSA integrates the outputs from three annotation tools ([ANNOVAR](#), [SnpEff](#) and [VEP](#)) for [RefSeq](#) and [Ensembl](#) GENCODE gene models, plus ANNOVAR outputs using [UCSC](#) knowGene model, and provides a summary of variant consequences from the seven annotation results. To further speed up the process for large-scale WGS studies, we have pre-computed annotations for all potential human SNVs (a total of 8,584,031,106) based on human reference hg19 and use it as a local database (in the [resources/precomputed](#) folder). For SNV-centric resources, WGSA integrated 13 functional prediction scores (CADD, FATHMM-MKL, Funseq, Funseq2, RegulomeDB, DANN, fitCons x 4, genoCanyon, Eigen, Eigen-PC), 8 conservation scores (GERP++, PhyloP x 3, phastCons x 3, SyPhy), allele frequencies from 4 large-scale re-sequencing studies (1000G, EP6500, ExAC, UK10K), variants in 4 disease related databases (ClinVar, COSMIC, GWAS_catalog, GRASP2), among others (see [list of resources](#)). For regulatory region-centric resources, WGSA integrated cell type specific transcription factor binding sites, DNase1 hypersensitivity regions and chromosome activity predictions from multiple epigenomics projects (see [list of resources](#)). WGSA also contains rich functional annotations for non-synonymous SNVs and genes from our [dbNSFP](#) database.

WGSA is provided both as an Amazon Machine Image (AMI) ready to run out-of-the-box and a downloadable version. WGSA and its resources are freely available for academic usage. Licenses are required for non-academic usage some of the resources, such as [ANNOVAR](#), [Polyphen2](#), [CADD](#), [DANN](#) and [VEST3](#) (in dbNSFP). WGSA does not grant the non-academic usage of those resources, so please contact the original content provider for that purpose.

You can sign up for [our email list](#) for future release announcements.

Please report any bug you found or any suggestions or comments to xmliu.uth@gmail.com or xiaoming.liu@uth.tmc.edu.

PredictSNP2



Unified platform for prediction of SNP effect in distinct genomic regions



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Job ID:

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INPUT

[Load example data](#)

[Show example results](#)

Genome assembly :  Format :

Insert the list of variants in one of the supported [formats](#) :

18,21118528,G,C

Load file : No file selected.

REFERENCE

Bendl, J., Musil, M., Stourac, J., Zendulka, J., Damborsky, J., Brezovsky, J., 2016: PredictSNP2: A unified platform for accurately evaluating SNP effects by exploiting the different characteristics of variants in distinct genomic regions. *PLOS Computational Biology* 12: e1004962.



USER STATISTICS

- Number of visitors: 5802
- Number of jobs: 8713

PredictSNP2: A Unified Platform for Accurately Evaluating SNP Effects by Exploiting the Different Characteristics of Variants in Distinct Genomic Regions

Jaroslav Bendl , Miloš Musil , Jan Štourač , Jaroslav Zendulka, Jiří Damborský , Jan Brezovský 

PredictSNP2



PREDICTSNP²

Unified platform for prediction of SNP effect in distinct genomic regions



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Job ID:

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JOB INFORMATION

ID: 10bwzv

Title: kurs_vcf

✓ Calculation successful, number of analyzed variants exceeded threshold for full table visualization. Only the 50 variants predicted as the most deleterious according to the user-selected tool are shown in the result table (box *Prioritize by tool*). The selection can be narrowed down to the specific category of variants, such as regulatory, splicing, missense, synonymous and nonsense subsets (box *Filter by category*).

RESULTS

RESULTS			neutral	deleterious	unknown	XX % expected accuracy		annotation										
Input			Prediction tools							Databases								
Variant	Region	Region function	PredictSNP2	CADD	DANN	FATHMM	FunSeq2	GWAVA	dbSNP	GenBank	Clinvar	OMIM	Regulome	HaploReg	UCSC	Ensembl	PredictSNP1	
2:166848563,C→G	exonic	nonsynonymous	87 %	52 %	60 %	83 %	61 %	51 %										
2:166903480,G→A	exonic	nonsynonymous	87 %	52 %	77 %	68 %	61 %	51 %										
2:166894356,C→T	exonic	nonsynonymous	87 %	52 %	70 %	80 %	61 %	50 %										
2:166198975,G→A	exonic	nonsynonymous	87 %	84 %	77 %	83 %	61 %	51 %										
12:52082568,G→A	exonic	nonsynonymous	87 %	84 %	72 %	83 %	62 %	50 %										
12:52159534,T→A	exonic	nonsynonymous	87 %	84 %	60 %	76 %	62 %	51 %										
9:130438189,G→A	exonic	nonsynonymous	87 %	84 %	77 %	83 %	62 %	51 %										
X:18606157,G→A	exonic	nonsynonymous	87 %	82 %	70 %	83 %	61 %	51 %										
9:130425622,C→T	exonic	nonsynonymous	87 %	84 %	77 %	56 %	62 %	50 %										
9:130444768,G→A	exonic	nonsynonymous	87 %	84 %	71 %	83 %	62 %	50 %										
Filter by category : All			Prioritize by tool : PredictSNP2			Genome assembly : GRCh37/hg19			Send to PredictSNP1									

Filter by category:

Prioritize by tool:

Genome assembly:

[Send to PredictSNP1](#)

PredictSNP2: A Unified Platform for Accurately Evaluating SNP Effects by Exploiting the Different Characteristics of Variants in Distinct Genomic Regions

Jaroslav Bendl , Miloš Musil , Jan Štourač , Jaroslav Zendulka, Jiří Damborský , Jan Brezovský

... Looking forward

<i>Day 1</i>
Genome Biology in 2017
Bioinformatics Tools in Epigenetics
Pathogenicity Assessment of DNA Sequence Mutations
Introduction to Galaxy
Massive Data Sources
Practical



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